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A Panel of Serum Biomarkers (GastroPanel®) in Non-invasive Diagnosis of Atrophic Gastritis. Systematic Review and Meta-analysis

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Abstract. Background/Aim: To meet the increasing demand of non-invasive tests for screening of gastric cancer (GC) risk, biomarker panel (GastroPanel®) (GP) was designed by Biohit Oyj as the first serological test for stomach health. The aim of the present study was to perform a systematic review and meta-analysis of all studies on GP in diagnosis of atrophic gastritis (AG). Materials and Methods: Studies were eligible, if i) GP was used to diagnose biopsy-confirmed AG of the corpus (AGC) and/or antrum (AGA) and ii) exact numbers were available to enable calculating sensitivity (SE) and specificity (SP). Comprehensive Meta-Analysis software was used with maximum likelihood meta-regression (R^2 analog). Effect size estimates (SE; SP, 95% confidence interval (CI)) were tested for homogeneity with Cochran's Q and I^2 statistics. Potential publication bias was estimated by funnel plot statistics. Results: Altogether, 27 studies were eligible comprising of 8,654 patients from different geographic regions. Significant heterogeneity between studies reporting AGC (n=27) or AGA (n=13) warranted random effects (RE) model for summary statistics. GP performs better in diagnosing AGC than AGA with 70.2% vs. 51.6% pooled SE and 93.9% vs. 84.1% pooled SP, respectively. Limited number of studies erodes the Q test's power to detect true heterogeneity in meta-analysis stratified by geographic study origin. Few hypothetical missing studies had only marginal effect on pooled estimates of SE and SP. Conclusion: This first

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Key Words: GastroPanel®, meta-analysis, sensitivity, specificity, biomarkers, pepsinogen I, pepsinogen II, gastrin-17, Helicobacter pylori, stomach mucosa, antrum, corpus, atrophic gastritis, meta-regression, gastric cancer, risk.

meta-analysis of GP literature corroborates the statement of international experts, advocating GP in diagnosis and screening of AG. Due to its high specificity for both AGA and AGC, GastroPanel® is truly a test for stomach health.

Gastric cancer (GC) remains the fifth most common malignancy worldwide with almost one million new cases and far over 700,000 annual deaths (1). This uniformly poor outcome of GC is due to the fact that, once diagnosed, the disease has already progressed beyond reach of curative therapy in most cases (2, 3). To improve the ominous disease outcome due to delayed diagnosis, novel diagnostic measures are urgently needed to allow early detection of GC (4).

Population-based screening by (invasive) endoscopy for early GC and its precursors is not feasible except perhaps in Japan (5). Atrophic gastritis (AG) and its causative etiological agent *Helicobacter pylori* (HP) are well-established precursors of non-cardiac GC (6-9). Therefore, non-invasive diagnostic tests for detection of AG and HP are promising tools for systematic screening of GC risk groups (5, 10-16). For some time, serum pepsinogen (PG) tests have been used for this purpose (5, 15); however, their impact on global GC mortality has been modest (17-19).

To meet the increasing demand, the GastroPanel® test (hereafter referred as GP) was designed in the late 1990's by Biohit Oyj (Helsinki, Finland) representing the first non-invasive diagnostic test for stomach health (13, 20, 21). This ELISA-based biomarker panel includes 3 markers of mucosal atrophy (PGI and PGII for the corpus; G-17 for the antrum), combined with HP IgG antibody assay (13, 16, 22). During the past decade, GP has been tested in different settings, mostly in diagnosis of symptomatic (dyspeptic) subjects (10, 13, 23-30). As emphasized (20-22), GP is not a test for invasive GC but designed for screening of the subjects at risk for GC, *i.e.* those with HP infection and AG. Until now, however, GP has been less extensively validated in population-based screening of such risk groups (11, 12, 30-33).

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The results on GP's sensitivity (SE) and specificity (SP) in diagnosis of AG have been variable (10-13, 23-34). There are several potential sources of bias affecting GP performance in different settings (22, 25). Apart from the different cut-off values used for PGI, PGII, G-17 and HP IgG in studies from different geographic regions, the prime culprit for the heterogeneity between the published studies is the misclassification bias or sampling error of gastric biopsies (4, 16, 22, 35-37). This is closely related to the use of appropriate histological end points in calculating the performance indicators of the GP test (10, 22, 30, 37).

We performed a systematic review and meta-analysis of the studies reporting the use of GP test in diagnosis of AG, covering all reports published until May 2016, with no restrictions to the language or geographic origin. The main objective was to estimate the SE and SP of the GP biomarkers in diagnosis of AGA (AG of the antrum) and AGC (AG of the corpus), separately. The maximum likelihood (ML) meta-regression was used to estimate the proportion of total between-study variance explained by the covariates in the model.

Materials and Methods

Data abstraction. We identified eligible studies by searching MEDLINE and reference lists from original articles, book chapters, reviews and congress abstracts until May 2016. No language limitations were imposed. The search terms included: GastroPanel, atrophic gastritis, biomarkers, pepsinogens, gastrin-17, Helicobacter pylori, stomach, gastric and human. All publications appeared in peer-reviewed journals, as well as published congress abstracts were eligible, irrespective of the study design (i.e., clinical or a screening setting). Most importantly, the eligible reports had to include GP test (Biohit Oyj, Helsinki, Finland) as the diagnostic tool. Thus, all studies using either pepsinogens or G-17 as stand-alone markers were excluded from this analysis as subjected to comprehensive meta-analysis previously (19). To be eligible, the report had to contain the exact numbers of gastric biopsies analyzed, as well as the numbers of AGC and/or AGA, examined by the GP biomarkers, pepsinogens and G-17, respectively (10, 22, 30). If not directly specified, the report had to be complete enough to enable the assessors to calculate these numbers from the results.

Using the above search terms, a database of 5,093 reports could be built by Reference Manager (Professional version 12.0.3.) covering the years 1936 through 2016. For the present meta-analysis, a total of 27 studies were determined eligible, fulfilling all the defined criteria. The absolute vast majority of the ineligible studies was irrelevant in type and appeared in the search due to a matching search term but otherwise lacking the necessary key identifiers needed for the meta-analysis. Another large category consists of studies where pepsinogens and/or G-17 were used as stand-alone markers in diagnosis of AG (19), without using the GP test.

Data extracted for meta-analysis. From the summaries and/or body texts of each eligible study, we extracted the following data: sample size (number of patients studied), topography of AGs (AGA or

AGC), number of AGA and AGC, geographic origin of the study, authors and publication year. To calculate the SE and SP for GP, we extracted the precise numbers of TP (true-positive), FN (false-negative), FP (false-positive) and TN (true-negative) (Table I). This list of items represents a modification of the PRISMA statements for systematic reviews and meta-analysis (38), adopted specifically for this purpose, where the effect size represents the SE and SP recorded for each original study. The meta-analysis file was arranged using AG as the study variable, whereby AGA and AGC represent subgroups within the study.

The authors defined GP performance indicators separately for PGI and G-17 used to diagnose AGC and AGA, respectively (10, 22, 30). In the next step, the authors identified from each eligible study the cut-off values for PGI and G-17 positivity (Table I). According to their geographic origin, the studies were stratified into the following categories: Central and South America, Europe, China or Taiwan and Other Asia. No studies were available from North America or from Africa/Middle East. At study level, the country of origin was classified into one of three categories, according to the incidence of GC; high-, intermediate- and low-incidence country. Publication year was arbitrarily divided into two categories; before year 2008 and after 2008, for older and more recent study, respectively.

Statistical analysis. A specific software, Comprehensive Meta Analysis[™] (Version 3.3.070; Biostat Inc., Englewood, NJ, USA), was used to perform the meta-analysis and meta-regression. The software calculates the SE and SP (and their logit event rate, standard error and variance) automatically when the numbers for TP/(TP+FN) and TN/(FP+TN) from the original data are entered as the events and the sample size, respectively, into the software's effect size data entry. To assess the overall heterogeneity in effect size estimates between the different studies, Cochran's Q homogeneity p-value, as well as I² statistics (for percentage of variation) were used (39). To explore the eventual publication bias, funnel plots were drawn by plotting the logit event rates by their precision (1/standard error) (40), evaluated for asymmetry using the following statistics: i) Begg and Mazumdar rank correlation (41), ii) Egger's test of the intercept (regression) (42) and iii) Duval and Tweedie's "trim and fill" method (43), which imputes the results that are hypothetically missing due to the publication bias.

To assess the variation in event rates due to the differences between individual studies, we evaluated the study characteristics using stratified meta-analysis and maximum likelihood (ML) meta-regression separately for both study subgroups (AGA, AGC) and for both effect size estimates (SE and SP). Meta-regression formally compares the differences in SE and SP across the selected study-level covariates and estimates the among-study variance (44). In meta-regression, different models were tested using the following study-level covariates in the model (collectively or as individual combinations): country, geographic origin, GC incidence (high-, intermediate-, low-), biomarker cut-off values, cohort size and time of publication (early, recent). In the new version of the software, the ML meta-regression estimates the proportion of total between-study variance explained by the set of covariates in the model, expressed as the R² analog (44).

Sensitivity analysis was performed to assess the influence of each individual study on the strength and stability of the meta-analytical results, using the one-by-one study removal and evaluated by descriptively comparing the magnitude and precision of the random-effects summary event rates.

Table I. Studies reporting GastroPanel sensitivity and specificity in diagnosing biopsy-confirmed atrophic gastritis of the corpus (AGC) and/or antrum (AGA).

Sample			Sensitivi	ty	,	Specifici	ty	Country	Authors	Year	Ref No
size	topography	TP	FN	Sensitivity (95%CI)	FP	TN	Specificity (95%CI)				No
104	AGC	32	3	91.4 (76.9-98.2)1	8	61	88.4 (78.4-94.9)	Italy	Zagari et al.	2002	33
	AGA	NA	NA		NA	NA					
13	AGC	2	2	50.0 (6.8-93.2) ⁶	0	6	100 (66.4-100)	Italy	DiMario et al.	2003	45
	AGA	NA	NA		NA	NA					
404	AGC	36	9	80.0 (65.4-90.4) ²	7	352	98.1 (96.0-99.2)	Finland	Väänänen et al.	2003	13
	AGA	2	2	50.0 (6.8-93.2) ³	5	180	97.3 (93.8-99.1)				
55	AGC	6	0	$100 (54.1-100)^4$	4	45	91.8 (80.4-97.7)	Poland	Hartleb <i>et al</i> .	2004	46
	AGA	7	4	63.6 (30.8-89.1)5	4	40	90.9 (78.3-97.5)				
178	AGC	46	16	$74.2 (61.5-84.5)^6$	17	99	85.3 (77.6-91.2)	Russia	Pasechnikov et al.	2004	47
	AGA	105	30	77.8 (69.8-84.5) ⁵	12	31	72.1 (56.3-84.7)				
109*	AGC	28	8	77.8 (60.8-89.9)6	3	70	95.9 (88.5-99.1)	Russia	Pasechnikov et al.	2005	48
	AGA	25	17	59.5 (43.3-74.4)5	9	58	86.6 (76.0-93.7)				
175	AGC	16	5	$76.2 (52.8-91.8)^6$	4	150	97.4 (93.5-99.3)	Italy	Cavallaro et al.	2004	23
	AGA	NA	NA		NA	NA					
50	AGC	9	3	$75.0 (42.8-94.5)^6$	0	38	100 (97.5-100)	France	De Korwin <i>et al</i> .	2004	24
	AGA	NA	NA		NA	NA					
287	AGC	38	22	63.3 (49.9-75.4) ⁶	16	211	93.0 (88.8-95.9)	Italy	Germanà et al.	2005	18
	AGA	NA	NA		NA	NA					
94	AGC	7	3	70.0 (34.8.93.3) ⁶	4	80	95.2 (88.3-98.7)	Italy	Nardone et al.	2005	29
	AGA	11	9	55.0 (31.5-76.9) ⁹	4	70	94.6 (86.7-98.5)				
180^	AGC	2	12	14.3 (1.8-42.8)6	0	145	100 (97.5-100)	Mexico	Graham et al.	2006	49
	AGA	20	20	50.0 (33.8-66.2)8	50	69	58.5 (48.6-68.0)				
56	AGC	7	1	87.5 (47.3-99.7) ⁶	0	48	100 (92.6-100)	Spain	Valle et al.	2007	50
	AGA	NA	NA		NA	NA					
71	AGC	5	2	71.4 (29.0-96.2) ⁶	2	13	86.7 (59.5-98.3)	Russia	Reshetnikov et al.	2008	51
	AGA	3	11	21.4 (4.7-50.8) ¹⁰	1	12	92.3 (64.0-99.8)				
976	AGC	44	18	71.0 (58.1-81.8) ^{5,6}	20	894	97.8 (96.6-98.7)	Sweden	Storskrubb et al.	2008	31
	AGA	NA	NA	460 (400 740)2	NA	NA	07.2 (02.2 00.2)			•	
162	AGC	6	7	46.2 (19.2-74.9) ²	4	145	97.3 (93.3-99.3)	Japan	Iijima <i>et al</i> .	2009	26
2.12#	AGA	2	11	$15.4 (1.9-45.4)^7$	5	144	96.6 (92.3-98.9)			•	
313#	AGC	3	3	100 (29.2-100)6	9	301	97.1 (94.6-98.7)	Italy	Guariso et al.	2009	52
00	AGA	NA	NA	00.0 (20.4.00.5)2	NA	NA	07 ((01 7 00 7)	T: 1 1	36 11	2000	
89	AGC	4	1	$80.0 (28.4-99.5)^2$	2	82	97.6 (91.7-99.7)	Finland	Mecklin et al.	2009	53
100	AGA	NA	NA	77.2 ((2.2.00.5)2	NA	NA	040 (02 1 07 0)	T. 1		2010	20
400	AGC	34	10	$77.3 (62.2-88.5)^2$	18	338	94.9 (92.1-97.0)	Italy	Lombardo et al.	2010	28
200	AGA	NA	NA	24 6 (17 2 55 7)6	NA	NA	00.0 (06.1.00.0)	0	D '4 1	2011	27
208	AGC	9	17	34.6 (17.2-55.7)6	2	180	98.9 (96.1-99.9)	Germany	Peitz et al.	2011	27
47**	AGA	32	83	27.8 (19.9-37.0)8	85	8	8.6 (3.8-16.2)	T4 - 1	M:1	2014	E 1
4/**	AGC	23	7	76.7 (57.7-90.1) ⁶	13	1	5.9 (0.5-28.7)	Italy	Masci et al.	2014	54
202	AGA	NA 24	NA 7	77.4 (62.7-92.1)11	NA 20	NA 54	72 6 (62 4 92 9)	Luca	Maiadi Valoniioni at	2014	55
283	AGC AGA	24 NA	/	77.4 (02.7-92.1)**	20	NA	72.6 (62.4-82.8)	Iran	Nejadi-Kelarijani et	2014	55
85	AGC	5	5	50.0 (18.7-81.3)6	15	60	90.0 (60.2.94.4)	Cnain	McNicholl et al.	2014	25
83				30.0 (16.7-61.3)			80.0 (69.2-84.4)	Spain	Michignon et at.	2014	23
100	AGA	NA 12	NA 7	65.0 (40.8-84.6) ⁶	NA	NA 79	00 0 (02 2 100)	Dussia	Ivoahlrin at al	2015	56
100	AGC AGA	13 54	7 5	91.5 (81.3-97.2) ⁵	1 0	41	98.8 (93.2-100) 100 (91.4-100)	Russia	Ivashkin et al.	2013	56
249	AGC	29	9	76.3 (59.8-88.6) ²	22	189	89.6 (84.6-93.3)	Italy	Goni et al.	2015	57
249				/0.5 (39.6-66.0)2			69.0 (64.0-95.5)	itary	Goiii ei ai.	2013	31
100	AGA	NA	NA 7	53.3 (26.6-78.7) ²	NA	NA 38	95.0 (83.1-99.4)	China	Gou et al.	2015	50
190	AGC	8 23	7 7	76.7 (57.7-90.1) ⁷	2	38 32	` /	China	Gou et at.	2015	58
2 050	AGA			81.3 (63.6-92.8) ²	8		80.0 (64.4-90.9)	Toirror	I ag at al	2015	50
2.858	AGC	26	6	` /_	21	88 87	80.7 (72.1-87.7)	Taiwan	Lee et al.	2015	59
019	AGA	11	7 11	61.1 (35.7-82.7) ⁷ 69.4 (51.9-83.7) ²	36 16	87 208	70.7 (61.9-78.6)	Dussia	Domar at al	2016	40
918	AGC	25	11	'	16	208	92.9 (88.7-95.9)	Russia	Roman et al.	2016	60
	AGA	10	28	26.3 (13.4-43.1) ⁷	35	189	84.4 (78.9-88.9)				

AG, Atrophic gastritis; AGC, atrophic gastritis in corpus; AGA, atrophic gastritis in antrum; TP, true-positive; FN, false-negative; FP, false-positive; TN, true-negative; CI, confidence interval; NA, data not available in the study; 1 G-17s, PGI and HPAb combined; 2 PGI (<30 µg/l); 3 G-17s (<3.0 pmol/l) + HPAb+(>30 EIU); 4 PGI (<50 µg/l); 5 G-17s (<5.0 pmol/l); *AGA and AGC only, Cases with multifocal AG were excluded from the analysis; 6 PGI (<25 µg/l); 7 G-17b (<1 pmol/l); 8 G-17b (<5 pmol/l) **All 47 cases represent antral or multifocal AG with no non-AG cases included; 6 180 subjects, of whom GastroPanel results available from 159; 9 G-17b (<2.5 pmol/l);); 10 G-17b (<2 pmol/l); $^{#}$ study subjects were children; 11 PGI <80 µg/l.

A Pooled Sensitivity (95%CI)

/bdel_	Study name	Subgroup within study	Statistics for each study								Event rate and 95% CI			
				Event rate	Lower	Upper limit	Z-Value	p-Value	Total					
	Väänänen et al	AGA	2003	0.500	0.123	0.877	0.000	1.000	2/4	1	1	1 -	-	_
	Hartlebet al.	AGA	2004	0.636	0.339	0.857	0.893	0.372	7/11	- 1	- 1	- 1		_
	Pasechnikovet al	AGA	2004	0.778	0.700	0.840	6.051	0.000	105 / 135	- 1	- 1	- 1		
	Pasechnikovet al	AGA	2005	0.995	0.443	0.731	1.227	0.220	25/42	- 1	- 1	- 1	+=-	
	Nardone et al	AGA	2005	0.550	0.336	0.747	0.446	0.665	11/20	- 1	- 1	- 1	_	-
	Graham et al.	AGA	2006	0.500	0.350	0.650	0.000	1.000	20/40	- 1	- 1	- 1	-	
	Reshetnikovet ala.	AGA	2008	0.214	0.071	0.494	-1.995	0.046	3/14	- 1	- 1	-	$\boldsymbol{\vdash}$	
	lijima et al.	AGA	2009	0.154	0.039	0.451	-2218	0.027	2/13	- 1	- 1	- =		
	Petzetal	AGA	2011	0.278	0.204	0.367	-4,580	0.000	32/115	- 1	- 1		■-	
	Ivashkin et al.	AGA	2015	0.915	0.812	0.964	5.090	0.000	54/59	- 1	- 1	- 1		-
	Couetal	AGA	2015	0.767	0.585	0.884	2.756	0.006	23/30	- 1	- 1	- 1		-
	Lee et al.	AGA	2016	0.611	0.379	0.802	0.935	0.350	11/18	- 1	- 1			-
	Roman et al	AGA	2016	0.263	0.148	0.424	-2.795	0.005	10/38	- 1	- 1	1 4	-	
Fixed				0.548	0.499	0.596	1.915	0.066	305/539	- 1	- 1		•	
ndom				0.538	0.383	0.687	0.477	0.634	305/539			1	-	

B Pooled Specificity (95%CI)

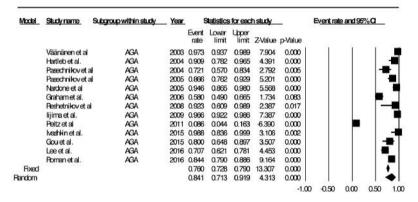


Figure 1. Pooled sensitivity and specificity of GastroPanel in diagnosis of AG of the antrum.

Results

Eligible studies. A total of 27 studies were considered eligible for the present analysis (13, 18, 23,-31, 33, 45-60) comprising of 8,654 patients analyzed by the GP test. Included are both small series (45, 54) and larger cohorts comprising up to 2,858 analyzed patients (31, 59, 60) (Table I). The studies include series where GastroPanel biomarkers were tested in diagnosing either AGA or AGC or both. In 14/27 reports, GP results on AGA are missing (18, 23-25, 28, 31, 33, 45, 50, 52-55, 57). As to the geographic origin of the studies, there were no studies published in North America and in Africa/Middle East.

Analytical results

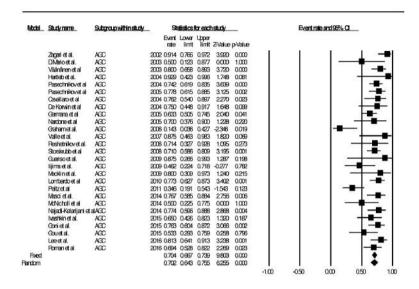
Pooled estimates of sensitivity and specificity. The pooled estimates of sensitivity and specificity of the GP test in diagnosis of AG were analyzed separately for the AG subgroups: AGA and AGC. Starting from AGA (n=13), the crude sensitivity (305=TP/539=TP+FN) translates to the pooled sensitivity of 0.548 (95% confidence interval (CI)=0.499-0.595), using the fixed effects (FE) model, and

0.538 (95%CI=0.383-0.687), using the random effects (RE) model (Figure 1A). Homogeneity (Cochran's Q)=107.525; I^2 =88.840; and p for homogeneity p=0.0001. For specificity, the crude figures (961=TN/1215=FP+TN) decode to a pooled specificity of 0.760 (95%CI=0.728-0.790) with the FE model and 0.841 (95%CI=0.713-0.919) with the RE model (Figure 1B). Homogeneity (Cochran's Q)=200.372; I^2 =94.011; and p for homogeneity p=0.0001.

Figure 2 depicts the meta-analytical results of pooled sensitivity and specificity for GP in diagnosis of AGC. The crude sensitivity (487/685) translates to pooled sensitivity of 0.704 (95%CI=0.667-0.739) with the FE model and 0.702 (95%CI=0.643-0.775) using the RE model (Figure 2A). Cochran's Q=52.807; I^2 =50.765; and p for homogeneity p=0.001. The crude specificity (3,946/4,171) corresponds to the pooled specificity of 0.924 (95%CI=0.914-0.934) using the FE model and 0.939 (95%CI=0.910-0.960) with the RE model (Figure 2B). Cochran's Q=196.475; I^2 =86.767; and p=0.0001.

Table II summarizes the meta-analytical results for GP sensitivity and specificity in diagnosis of AGC, stratified by the geographic origin of the study. As to the test sensitivity,

A Pooled Sensitivity (95%CI)



B Pooled Specificity (95%CI)

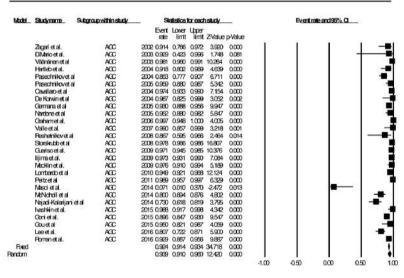


Figure 2. Pooled sensitivity and specificity of GastroPanel in diagnosis of atrophic gastritis of the corpus (AGC).

heterogeneity is significant in the overall comparison between strata (RE model, p=0.017). Using the RE model, studies from Europe give the highest pooled estimates of sensitivity (72.1%). As to the test specificity, there is a significant heterogeneity between the studies (n=26) from China/Taiwan, Europe and Other Asia, with p=0.048, p=0.0001 and p=0.0001, respectively. However, heterogeneity is not significant in the overall comparison between strata (RE model, p=0.199). Using the RE model, studies from Europe

give the by far highest pooled estimates of specificity (0.943, *i.e.* 94.3%), followed by those from Other Asia (89.8%).

Table III shows the meta-analytical results for GP sensitivity and specificity in diagnosis of AGA, stratified by the geographic origin of the study. As to the test sensitivity, heterogeneity is not significant in the overall comparison between strata (RE model, p=0.483). Using the RE model, studies from China/Taiwan give the highest pooled estimates of sensitivity (69.6%), followed by those from Europe (54.5%). Regarding the test specificity,

Table II. Pooled sensitivity and specificity of GastroPanel for atrophic gastritis of the corpus (AGC) in stratified meta-analysis by geographic study origin.

Geographic origin of study	No. of studies	Events	Sample size	effe	stimates of ect size model)	effec	timates of et size model)	Heterogeneity (Cochran's Q)	**I-squared (I ²)	Heterogeneity (p-value)
				Point Estimate	95%CI	Point Estimate	e 95%CI			
Sensitivity										
China/Taiwan	2	34^{1}	47^{2}	0.709	0.555-0.826	0.701	0.496-0.848	3.756	73.374	p=0.053
Europe	22	421^{1}	580^{2}	0.715	0.675-0.752	0.721	0.663-0.773	32.519	35.423	p=0.052
Other Asia	2	30^{1}	442	0.671	0.512-0.799	0.656	0.444-0.820	3.890	74.293	p=0.049
S/C America	1	2^{1}	14^{2}	0.143	0.036-0.427	0.143	0.030-0.474	0.000	0.000	p=1.000
Summary Total within (FE) Total between (FE) Total between (RE)	27	4871	685 ²	0.704	0.677-0.739	0.615	0.418-0.780	52.807 40.165 12.643 10.136	50.765	p=0.001 p=0.015 p=0.005 p=0.017
Specificity										
China/Taiwan	2	126^{3}	149^{4}	0.830	0.757-0.885	0.884	0.642-0.970	3.904	74.385	p=0.048
Europe	22	3.476^{3}	3.654^4	0.938	0.929-0.947	0.943	0.913-0.963	126.345	83.379	p = 0.0001
Other Asia	2	199^{3}	2234	0.823	0.747-0.880	0.898	0.686-0.972	20.729	95.176	p = 0.0001
S/C America	1	145^{3}	145 ⁴	0.997	0.948-1.000	0.997	0.914-1.000	0.000	0.000	p=1.000
Summary Total within (FE) Total between (FE) Total between (RE)	27	3.9463	4.1714	0.924	0.914-0.934	0.936	0.846-0.975	196.475 150.977 45.498 4.654	86.767	p=0.0001 p=0.0001 p=0.0001 p=0.199

^{**}Only calculated for fixed effects model: FE, fixed effects model; RE, random effects model; CI, confidence interval; ¹Events=TP; ²Sample size=TP+FN; ³Events=TN; ⁴Sample size= FP+TN; S/C, South and Central.

heterogeneity is not significant in the overall comparison between strata (RE model, p=0.532). Excluding the regions with only one study, the studies from Europe give the highest pooled estimates of specificity (RE model) (86.0%), followed by those from China/Taiwan (75.6%).

Meta-regression. In ML meta-regression for GP sensitivity in diagnosis of AGA, the model contained the following covariates: GC incidence (high-, intermediate-, low-), G-17 cut-off values, cohort size, study timing, resulted in an R² analog of 0.610, i.e. 61.0% of the variance in true effects can be explained by these covariates. As to GP specificity for AGA, 95% of the variance (R² analog=0.950) could be explained by the model with the following covariates: geographic study origin, G-17 cut-off values and study timing. Changing the actual study year by the covariate early/recent, resulted in a model explaining the between-study variance in full (R² analog=1.00).

As to GP sensitivity for AGC, 96% of the variance (R² analog=0.960) in true effects could be explained by the model with the following covariates: geographic study origin, GC incidence (high-, intermediate-, low-) and PGI cut-off values. Adding the cohort size as the covariate,

resulted in a model explaining the between-study variance in full (R^2 analog=1.00). As to GP specificity for AGC, the best fitting model included the following covariates: geographic study origin, GC incidence (high-, intermediate-, low-), PGI cut-off values, early/recent study) and explained 81% of the variance (R^2 analog=0.810) in true effects.

Publication bias. Publication bias was analyzed using precision funnel plots and the test statistics (Figure 3). Among studies reporting GP sensitivity in AGA end point, there was no evidence for publication bias; Begg p=0.541, Egger's p=0.923, the Duval and Tweedie's trim and fill method identified no missing studies (RE, to left or right of mean) (Figure 3A). For studies reporting GP specificity for AGA, the same was true with Begg p=0.179 and Egger's p=0.216; the Duval and Tweedie's trim and fill method imputed 4 hypothetically missing studies with a substantial effect on pooled estimate (RE) of specificity (from 84.1% to 72.8%) (Figure 3B).

Among studies reporting GP sensitivity for the AGC end point, there was no evidence for publication bias analyzed by the Begg (p=0.851) or Egger's (p=0.988) tests. The Duval and Tweedie's trim and fill method (RE) identified 4 missing

Table III. Pooled sensitivity and specificity of GastroPanel for atrophic gastritis of the antrum (AGA) in stratified meta-analysis by geographic study origin.

Geographic origin of study	No. of studies	Events	Sample size	effe	stimates of ect size model)	effec	cimates of et size model)	Heterogeneity (Cochran's Q)	**I-squared (I ²)	Heterogeneity (p-value)
				Point Estimate	95%CI	Point Estimate	95%CI			
Sensitivity										
China/Taiwan	2	34^{1}	48^{2}	0.703	0.558-0.817	0.696	0.295-0.926	1.295	22.783	p = 0.062
Europe	9	249^{1}	438^{2}	0.543	0.488-0.597	0.545	0.348-0.729	95.411	91.615	p=0.0001
Other Asia	1	2^{1}	13^{2}	0.154	0.039-0.451	0.154	0.012-0.728	0.000	0.000	p=1.000
S/C America	1	20^{1}	40^{2}	0.500	0.350-0.650	0.500	0.090-0.910	0.000	0.000	p=1.000
Summary Total within (FE)	13	3051	539 ²	0.548	0.499-0.595	0.522	0.285-0.749	107.525 96.706	88.840	p=0.0001 p=0.0001
Total Between (FE)								10.818		p=0.013
Total Between (RE)								2.457		p=0.483
Specificity										
China/Taiwan	2	119^{3}	163^{4}	0.728	0.654-0.791	0.756	0.263-0.964	1.299	22.999	p=0.254
Europe	9	629^{3}	784^{4}	0.808	0.768-0.843	0.860	0.677-0.947	149.138	94.636	p=0.0001
Other Asia	1	1443	149^{4}	0.966	0.922-0.986	0.966	0.559-0.998	0.000	0.000	p=1.000
S/C America	1	69^{3}	119 ⁴	0.580	0.490-0.665	0.580	0.063-0.966	0.000	0.000	p=1.000
Summary	13	961 ³	1.2154	0.760	0.728-0.790	0.839	0.606-0.947	200.372	94.011	p=0.0001
Total within (FE)								150.436		p=0.0001
Total between (FE)								49.936		p=0.0001
Total between (RE)								2.200		p=0.532

^{**}Only calculated for fixed effects model: FE, fixed effects model; RE, random effects model; ¹Events=TP; CI, confidence interval; ²Sample size=TP+FN; ³Events=TN; ⁴Sample size= FP+TN; S/C, South and Central.

studies (left of mean) with a marginal effect on pooled estimate (RE) of sensitivity (from 70.2% to 68.1%) (Figure 3C). For studies reporting GP specificity for AGC, the same was true with Begg (p=0.851) and Egger's (p=0.229) tests, whereas the Duval and Tweedie's trim and fill (RE, left of mean) method imputed 5 hypothetically missing studies with only a slight effect on pooled estimates (RE) of specificity: from 93.9% (observed) to 92.5% (adjusted) (Figure 3D).

Sensitivity analysis. In general, the meta-analytical results seemed quite robust to all (n=27 AGC, n=13 AGA) one-byone study removals with little change in the magnitude and precision of the FE- and RE-pooled estimates of GP sensitivity and specificity. However, among studies reporting sensitivity for AGA, there are two influential studies (27, 47) that, if omitted, have an impact on pooled sensitivity estimates (FE) from 54.8% up to 63.% (27) and down to 46.8% (47), respectively. For studies reporting GP sensitivity in AGC, no influential studies were identified by either FE or RE models. As to GP specificity in AGA, two studies had a moderate impact on pooled estimates but only in FE model (17, 49); once removed, the pooled specificity estimate

improved from 76.0% to 79.6% and 80.0%, respectively. GP specificity was absolutely robust to all one-by-one study removals, with each study affecting the pooled estimates with less than 0.010, (i.e. <1% only), irrespective whether FE or RE model was used (data not in Tables).

Discussion

GastroPanel® test is a biomarker panel based on simultaneous analysis of PG-I, PG-II, amidated G-17 and HP IgG antibodies, designed to give information on both the structure and function of the stomach mucosa (13, 20-22). The added value of using this 4-biomarker combination instead of PGs and G-17 as stand-alone markers (14-16, 19) lies in the fact that this single test provides comprehensive information from the entire stomach, not restricted to either antrum or corpus only (10-13, 16, 20-22). GP results are interpreted by a specific software (GastroSoft®), classifying the test results into one of five categories, each with a specific biomarker profile (10-13, 16, 20-22, 30-32) and matching the diagnostic categories of the Updated Sydney System (USS) of classifying gastritis (35-37).

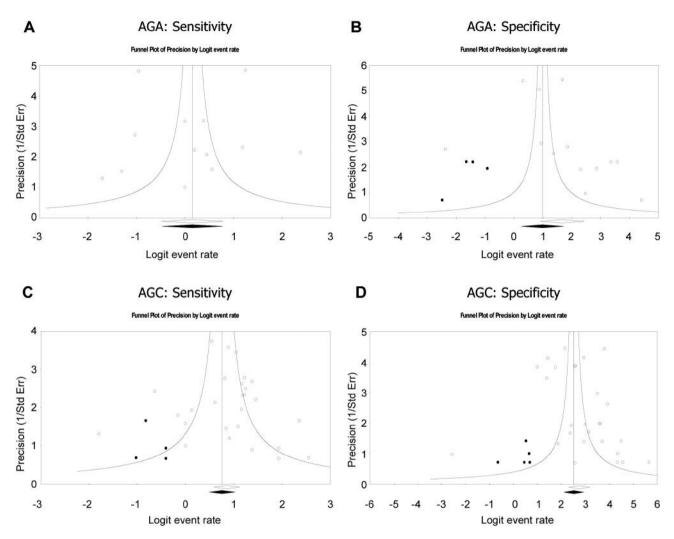


Figure 3. Publication bias of the GastroPanel studies for atrophic gastritis of the antrum (AGA) and atrophic gastritis of the corpus (AGC) end points estimated by precision funnel plots.

The experience on GastroPanel[®] test has been variable and, in part, conflicting, despite the recent consensus statement by an international panel of experts advocating its use in diagnosis and screening of AG (16). To cast further light on the potential causes of the between-study variance in the reported sensitivity and specificity of GP test, we performed a systematic review and meta-analysis covering all published studies where GP was used in diagnosis of AGA and AGC. Different GP biomarkers are used for AGA and AGC, G-17 and PGI, respectively (13, 16, 22, 30-32); AGA and AGC were treated as subgroups of AG in the present meta-analysis (Table I).

To assess the heterogeneity in meta-analysis is essential because the presence or absence of true heterogeneity (i.e.

between-study variance) directly affects the statistical model that should be applied to analyze the meta-data (39, 64-67). Using the Q test, introduced by Cochran (67), a nonsignificant homogeneity p-value justifies the adoption of a fixed-effects (FE) model, assuming that the estimated effect sizes only differ by sampling error (64). In contrast, significant p-values in the Q test indicate true heterogeneity, advocating the use of a random effects (RE) model that includes both within- and between-studies variance. The I^2 index measures the extent of true heterogeneity, interpreted as the percentage of the total between-study variance among the effect sizes (68). One of its definite advantages is that the I^2 indices obtained from meta-analyses with different numbers of studies and different effect metrics are directly comparable (64, 68).

There is little doubt that a marked heterogeneity exists between the studies reporting GP sensitivity and specificity in both AGA and AGC (Figures 1 and 2; see Results for the exact Q-, I²-index- and *p*-values). This advocates the use of the RE model to analyze all the summary statistics (39, 64-68). As shown by the forest plots for AGA and AGC (Figures 1 and 2), GP test performs better in diagnosing AGC than AGA. The difference is more marked in test sensitivity, with pooled (RE) estimates of 0.538 and 0.702 in diagnosis of AGA and AGC, respectively. The difference is less marked in test specificity for AGA and AGC, 0.841 and 0.939, respectively. Even more marked between-study heterogeneity was reported in a meta-analysis of the studies (n=12) using PGI and/or PGI/PGII as stand-alone markers for AGC, with SE varying between 5.8% and 98.6% and SP between 64.0% and 100% (19).

In practical terms, PGI biomarker of the GP detects AGC with 70.2% pooled sensitivity and 93.9% pooled specificity, whereas G-17 detects AGA with 53.8% pooled sensitivity and 84.1% pooled specificity. These meta-analytical results are not unexpected and the pooled estimates of GP sensitivity and specificity closely match those obtained in our own validation studies (13, 31, 32, 60). The reasons for this different GP performance in AGC and AGA are to be found in the different target-specificity and functional regulation of PGI and G-17 biomarkers (13, 22). As PGI is a product of gastric corpus, and the below-cut-off levels (30 μg/l) are only possible when the chief cells disappear as the result of mucosal atrophy, PGI is a highly sensitive and specific marker of AGC (10, 14-16, 22). G-17, in turn, is a specific biomarker of the antrum expressed by the G-cells that disappear in AG of the antrum (22, 48-50).

However, it is well-established that low levels of G-17b are not exclusively inherent to AGA but also result from high gastric acid output by the corpus (22, 69-71). On the other way round, G-17 is up-regulated (through a negative feedback loop) by low acid content of the corpus, caused by either i) AGC or ii) a prolonged use of proton pump inhibitor (PPI) medication (10, 14-16, 22, 69-71). By definition, any biomarker that is being regulated by more than one trigger cannot be a highly specific indicator of only one of them. In the case of fasting G-17b, the below-cut-off values can be due to either AGA or high acid output (22). Thus, for a distinction between these two potential causes of low G-17, G-17b measurement alone is not sufficient but it should be complemented by measuring G-17 levels after protein stimulation, i.e. G-17s, which is another component of the GP test (22). Failure to increase G-17s output after such a stimulation implicates the lack of G-cells and the presence of AGA (22, 69-71). Of the studies included in this meta-analysis, only four had included G-17s measurement in their GP analysis (13, 46-48). This precludes the possibility of making the distinction between AGA and high acid output as the cause of low G-17 levels (69-71). Failure to do so inevitably corrodes the sensitivity of G-17 as the marker of AGA, levelling off at 53.8% in the pooled (RE) analysis (Figure 1).

GC and its precursors (HP infection and AG) demonstrate a significant geographic variation in their global occurrence (1, 2, 10-13, 16, 19, 30-32). Accordingly, geographic origin of the GP studies clearly is a potential study-level covariate, here addressed by stratified meta-analysis. Interestingly, for AGA, the stratified meta-analysis using O test and I² index did not disclose a marked heterogeneity between the studies from the four geographic regions (Table III). The most likely explanation is the limited number of studies reporting AGA, with two of the regions only having one single study, thus limiting the power of Q test to disclose true heterogeneity (39, 64-67). As to AGC, the stratified meta-analysis confirmed substantial heterogeneity only for the sensitivity (Table II). For specificity, there was no significant betweenregion variance (p=0.199). The small number of studies from three of the regions (n=2, n=2 and n=1) is probably the most important limiting factor for adequate power of the Q test to detect true heterogeneity.

Finally, we performed the ML meta-regression to estimate the proportion of total between-study variance explained by the set of covariates in the model. In the new version of the meta-analysis software, this is expressed as the R² analog (61-63), which is analogous to the R² index commonly reported for the proportion of variance explained by covariates in the primary studies. In the primary studies, all observations are given the same weight, while, in meta-regression, each study is given a different weight (61). In our meta-regression, the following study-level covariates were included in the model: country, geographic origin, GC incidence (high-, intermediate-, low-), biomarker cut-off values, cohort size, year of publication, early/recent study.

For the AGA end-point, we could build-up a regression model that explained at beast 61% of the variance in true sensitivity of the GastroPanel[®] test. For test specificity, in turn, we could find a set of covariates that explained the true variance by 100%, *i.e.* R2 analogue was 1.0. This is in alignment with the view that G-17b is not a highly sensitive marker of AGA as it is affected by both AGA and high acid output (22, 69-71). This information was not available as a covariate in the present meta-analysis. However, this information is not necessary to correctly classify the antrum as normal (AGA-negative) by normal G-17b levels, implicating that the existing covariates are sufficient to fully explain the true between-study variance in GP specificity.

In AGC, the reverse is true, *i.e.* the existing covariates resulted in a model explaining the between-study variance of sensitivity in full but the true variance of specificity by 81% only. This suggests that the below-cut-off levels of PGI are sensitive indicators of AGC, whereas additional covariates are needed to classify gastric corpus healthy (AGC-negative). One such candidate is the presence of HP infection, as well as the

PGI/PGII ratio, which both are essential components of the full GastroPanel® test; however, not included as separate covariates in the present meta-analysis (because incompletely reported). According to our experience, PGI/PGII ratio is another powerful marker of AGC, equivalent or sometimes superior to PGI alone (11-13, 16, 22, 30-32, 60). It might well be that studies using PGI alone might misclassify a few cases of AGC, being responsible for the between-study variance in test specificity that is not being explained by the set of available covariates. Importantly, the GastroSoft® algorithm also takes into account G-17 while interpreteing the GP results; the complete biomarker profile of AGC includes i) low PGI, ii) low PGI/PGII ratio, as well as iii) increased G-17b (16, 22, 30-32, 69-71). These data were not available from the original studies for use as covariates in this analysis. Similarly, diagnosis of AGA is made by GastroSoft® only when both conditions: i) low G-17 and ii) HP Ab titer above the cut-off (30 EIU) are fulfilled (22, 69, 70). As said, G-17s is needed in confirming the antrum atrophy.

Taken together, the present meta-analysis disclosed significant between-study heterogeneity in reported sensitivity and specificity for both AG end points. GastroPanel® performs better in diagnosing AGC than AGA, with 70.2% vs. 53.8% pooled sensitivity and 93.9% vs. 84.1% pooled specificity, respectively. In meta-analysis, stratified by the geographic origin of the study, the between-region heterogeneity was less pronounced than anticipated on the basis of the highly divergent geographic variation in GC and its precursor lesions (10-13, 16, 19, 30-32). Undoubtedly, the limited number of studies per strata erodes the power of the Q test to detect true heterogeneity in this stratified analysis.

Of the potential sources of errors affecting GastroPanel® performance in different studies, most important is the misclassification bias of gastric biopsies (22, 35, 37). This can result from two possible sources: a) failure to identify the correct biopsy site on gastroscopy (e.g. patchy AG) and b) unsatisfactory intra- and inter-observer reproducibility of grading the AG (22, 35-37). The different potential of G-17 and PGI to diagnose AGA and AGC, respectively, is inherent to their different physiological regulation (22, 69-71). GastroPanel® is an entity of 4 stomach-specific biomarkers but the value of this diagnostic test is much more than the sum of components. GastroPanel® gives comprehensive information on the structure and function of the gastric mucosa and diagnosing AGA and AGC is just one part of the diagnostic range of this unique test. As suggested by the independent panel of international experts, GastroPanel® should be the firstline test in diagnosis and screening of AG (16). The present study, reporting the first formal meta-analysis of the GastroPanel® literature, clearly corroborates this statement; due to its high specificity in diagnosing both AGA and AGC, GastroPanel® is truly a test for stomach health (16, 22) with excellent longitudinal negative predictive value for GC (30).

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